www.jneurovirol.com

Endothelin and nitric oxide levels in cerebrospinal fluid of patients with multiple sclerosis

Livianna Speciale¹, Marina Sarasella¹, Stefania Ruzzante¹, Domenico Caputo², Roberta Mancuso¹, Maria Gaetana Calvo¹, Franca Rosa Guerini¹ and Pasquale Ferrante^{*,3}

¹Laboratory of Biology, Don C. Gnocchi Foundation, IRCCS, Milan, Italy; ²Multiple Sclerosis Unit, Don C. Gnocchi Foundation, IRCCS, Milan, Italy and ³Chair of Virology, Department of Preclinical Sciences, University of Milan, Italy

> In order to investigate the potential role of endothelins (ETs) and nitric oxide (NO) in the pathogenesis of multiple sclerosis (MS) we evaluated the levels of these vasoactive mediators in cerebrospinal fluid (CSF) of relapsing remitting MS patients and in a group of subjects with other neurological diseases (OND) and in a control group of subjects without neurological disease. Eighty patients affected from clinically diagnosed MS were selected, 44 of them were studied during an acute clinical attack and 36 in a stable phase. The OND group included 21 subjects affected by degenerative non inflammatory (n=9) and inflammatory (*n*=12) neurological disease while the control group included 22 subjects with cancer of the prostate (n=11) and with bladder disease (n=11). ET levels were significantly increased in CSF of relapsing remitting MS patients with an acute clinical attack in comparison with those in a stable phase, the OND group and the control group. Moreover significant differences were observed among the four groups with regard to the NO levels: MS patients in a stable and acute phase like OND group have high levels of NO compared to the control group. Since the blood-brain barrier index values did not differ significantly between the three groups, the data of this study suggest an important role for NO and ET in cerebral microcirculation in MS patients. Journal of NeuroVirology (2000) 6, S62-S66.

Keywords: endothelins; nitric oxide; multiple sclerosis; cerebrospinal fluid

Introduction

In multiple sclerosis (MS), demyelination is the result of a series of pathological events which include local oedema, perivascular infiltration and production of cytokines and neurotoxic substances. It has been recently suggested that various substances may be involved within the central nervous system (CNS) and among the other endothelins (ETs) (Levin, 1995; Rubaniy, 1992) and nitric oxide (NO) (Merril *et al*, 1993; Hooper *et al*, 1995) are a group of vasoactive mediators suspected of playing a leading role. Endothelins are produced by endothelial and microglial cells and astrocytes in CNS as neuro-transmitters (Harland et al, 1995) and are involved in CNS cerebrovascular disorders (Fujimori et al, 1990; Suzuki et al, 1990; Ehrenreich et al, 1992). Some reports indicate that Endothelin 1 probably acts as modulator of inflammatory status in CNS up-regulating the local expression of intercellular and vascular adhesion molecules in microvascular endothelial brain, implicating the peptide in the recruitment of blood cells at site of inflammation (Barone *et al*, 1995). ET also contribute to mitogenesis and proliferation of the glia and astrocytes (Ross and Snyder, 1990; Harland *et al*, 1995) and can participate in tissue regeneration (Yamada *et al*, 1995).

Also NO can influence cerebrovascular resistance and the permeability of the blood-brain barrier under pathological condition and is over produced by microglial cells during inflammation in the brain (Wong *et al*, 1996) becoming responsible for demyelinating process (Merril *et al*, 1993) death and loss of oligodendrocytes (Ikeda *et al*, 1995). Moreover low levels of NO lead to a decrease in myelin production from oligodendrocytes (Merril *et al*, 1993).

In this study the potential role of ETs and NO in MS pathogenesis has been investigated by measuring their level in cerebrospinal fluid (CSF) collected from relapsing remitting MS patients and from a group of patients with other neurological diseases (OND) and in a control group without neurological diseases. A significant

^{*}Correspondence: P Ferrante, Laboratory of Biology, Don C. Gnocchi Foundation, IRCCS, Via Capecelatro, 66, I-20148, Milan, Italy

increase of ETs and NO levels have been observed in the CSF of MS patients.

Results

The results of the measurement of ET levels in the CSF of the four groups of patients is exposed in the Figure 1A. ET levels (mean=11.0; s.d.=12.67 pg/ml) in CSF of acute relapsing remitting MS patients were significantly higher (P < 0.05) than those observed in stable relapsing remitting MS patients (mean=5.1; s.d.=9.13 pg/ml) and the OND group (mean=2.83; s.d.=4.3 pg/ml) (P < 0.05) and the control group (mean=1.5; s.d.=2.3) (Figure 1). The data obtained by measuring the NO levels in the CSF of the same patients are reported in Figure 1B. Significant differences in total nitrate/nitrite have been observed between acute relapsing remitting MS (mean=54.52; s.d.=23.9) and stable relapsing

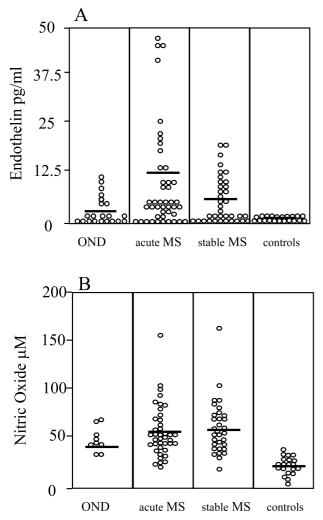


Figure 1 Distribution of Endothelin (A) and Nitric Oxide (B) levels in CSF from OND patients, acute and stable MS patients and not neurological controls.

remitting MS (mean=56.53; s.d.=27.6) versus the control group (mean=28.7; s.d.=8.1) and in the OND patients (mean=44.29; s.d.=11.0) versus the control group. No differences were observed in the two groups of MS patients.

In order to verify if the presence in the CSF of ET and/or NO could be due to damage of a BBB, the IgG intrathecal synthesis index and the Link IgG index have been evaluated in all the patients studied. The results obtained are shown in Table 1 in which it is possible to see that the value of BBB integrity does not differ among the three groups of the MS or OND patients while as expected the IgG index values are increased in MS patients but are within normal range in the OND cases.

Discussion

ETs have been suggested to act as neuropeptides in the CNS and to modulate neuronal activity (Giaid et al, 1991). In normal brain glial cells, including astrocytes and endothelial capillary cells, do not express endothelin-like immunoreactivity while the production of ETs is increased in presence of CNS diseases and injuries (Lee et al, 1990; Jiang et al, 1993). For instance reactive human astrocytes and macrophages express ET mRNA during viral CNS diseases, such as progressive multifocal leukoencephalopathy (PML) and subacute sclerosing panencephalitis (SSPE) (Ma et al, 1994). In the present study we report the presence of enhanced levels of ETs in CSF of acute relapsing remitting MS patients in comparison to those in a stable disease phase and to the OND patients. Moreover the elevated levels of nitric oxide in CSF of patients with MS provides further evidence for NO in immunopathogenesis of MS according to already published data (Giovannoni, 1998). However nitrite (NO_2^-) and nitrate (NO_3^{-}) as charged anions have difficulty to cross the blood-brain barrier and this suggests that a greater quantity of intrathecal NO is produced in MS patients (Felgenhauer *et al*, 1982).

Moreover we did not observe a higher frequency of BBB damage in acute MS patients, the increased ETs levels found in the CSF of these patients are probably due to a local synthesis by glia, neurons, astrocytes or infiltrating monocytes. Although the role of this peptide family is still not clearly defined

Table 1IgG synthesis index and blood-brain barrier levels inCSF of patients with acute MS, stable MS and OND reported asmean value and standard deviation.

OND	Acute MS	Stable MS
0.668 ± 0.86	0.811 ± 0.46	0.701 ± 0.22
5.16 ± 4.08	5.034 ± 1.92	5.191 ± 2.25
	0.668±0.86	0.668 ± 0.86 0.811 ± 0.46

it has been suggested that the increased synthesis of ET during CNS injury promotes the mitogenesis of glioma cells and astrocytes and may contribute to the reduction of the local blood flow (O'Brien et al, 1992; Bonte et al, 1993), and that, on the other hand, it could play a role in the extravasation and migration of peripheral blood lympho-monocytes through the BBB (MacCumber et al, 1990; Yamada et al, 1995; Kohzuki et al, 1995). It has also been suggested that ETs may participate in tissue regeneration since they are largely expressed from astrocytes in the areas surrounding necrotic lesions or demyelinating plaques in PML and SSPE (Yamada et al, 1995). Moreover these peptides have been shown to be capable of promoting the activation of astrocytes and microglial cells (Holzwarth *et al*, 1992; Lin and Chuang, 1992; Jiang *et al*, 1993; Stanimirovic et al, 1995) and therefore to induce the production of Transforming growth factor beta (TGF β) (Lindholm *et al*, 1992; Merril *et* al, 1993; Yamada et al, 1995), a cytokine that induces differentiation of the precursor of oligodendrocytes into myelin-producing cells in vitro (Eddleston et al, 1993; Gard et al, 1995).

Increased levels of NO have been observed in Parkinson, Alzheimer and in HIV-1-induced CNS diseases suggesting a neurotoxic activity of this neuro-transmitter molecule (Qureshi et al, 1995; Goodwin et al, 1995). NO causes injury to oligodendrocytes also if the adjacent astrocytes and microglial cells are not damaged or killed in vivo and in vitro (Prineas, 1985; Merril et al, 1993). Since the oligodendrocytes seem to be the target in MS pathogenesis it is possible that a lower concentration of NO lead to the damage of oligodendrocytes by alteration of myelin biochemical turnover (Merril and Benveniste, 1996). Our data show the presence of high levels of NO in CSF of MS and OND versus control group. The presence of NO in CSF of MS patients contribute to the hypothesis that the low levels of NO can mediate the blood-brainbarrier breakdown that occur in CNS inflammation.

On the whole the results of our study suggest that the higher ET levels observed in the CSF of the acute MS patients may be indicative of its potential as a marker of disease activity, while the presence of NO levels may be considered as an indicator of BBB dysfunction in CNS inflammation. The role of ETs and NO in the inflammatory process leading to MS pathogenesis requires further investigation and more evidence is needed to establish whether ETs play a role in triggering local reduction of blood flow in the CNS or if they are produced as consequence to damage to endothelial cells.

Materials and methods

Patients and controls

The CSF samples analysed in this study were collected from 80 MS patients (51 females and 29

males, mean age 38 years) with clinically definite relapsing remitting MS diagnosed according to the criteria of Poser *et al* (1993). MS patients with a new clinical acute attack and with a cerebral gadolinium-enhancing area evidenced by magnetic resonance imaging (MRI), performed on a 0.5 Tesla operating unit (General Electric MR MAX), were defined as acute, while those without a clinical relapse in the previous 6 months and without gadolinium-enhancing areas were classified as stable MS. According to the clinical observation and to the MRI results of the 80 RRMS patients 44 were in the acute phase whereas 36 were in the stable phase of disease.

The mean period of disease is 6.7 years for the RRMS in stable phase and 6 years for the patients in acute phase. None of the patients had received ACTH, corticosteroid or any other relevant pharmaceutical agent for at least 3 months, or any immunosuppressive agent for at least 6 months before CSF collection. Atypical dietary habit rich in nitrates was investigated and if present used as exclusion criteria.

A group of 21 patients suffering from other neurological diseases (OND) was studied as control. Ten of these were females and 11 males with a mean age of 35 years and a mean disease duration of 1.6 years. This group included nine patients with degenerative non inflammatory neurological disease (seven amyotrophic lateral sclerosis (ALS), one labyrinthine syndrome, one spastic paresis) and 12 with inflammatory neurological diseases (four encephalomyelitis, five pyramidal syndrome, 2 Beçhet, one cerebellar ataxia).

The non neurological control group included 22 subjects (21 males, one female) suffering from cancer of the prostate (n=11) and of bladder (n=11).

CSF routine laboratory analysis

For all the MS patients and the OND controls, routine analyses were performed on the CSF samples obtained by sterile lumbar puncture. Blood-brain barrier (BBB) integrity, the IgG intrathecal synthesis indexes and the presence of IgG oligorlonal bands (OB) were evaluated using standard procedures. The Link IgG index, (Lefvert and Link, 1985) the Tourtellotte IgG synthesis index (Tourtellotte et al, 1985) and BBB integrity value were calculated by measuring serum and CSF IgG and albumin levels, using the nephelometer system APS Beckman (Beckman Instruments, Inc., Galway, Ireland). Normal values were established as $\leq 0.7 \text{ mg/ml/day}$ for the Link IgG index, $\leq 3.3 \text{ mg/}$ dl/day for the Tourtellotte IgG synthesis and $\leqslant 5.5$ for the BBB integrity index.

The presence of CSF IgG OB was evaluated by sodium dodecylsulfate (SDS) acrylamide gel electrophoresis with silver staining, using the Phast System apparatus (Pharmacia Biotech, Uppsala, Sweden). Endothelin levels in the CSF were assaved using the Endothelin Immunoassay Kit (Cayman Chemical Company, Ann Arbor, MI, USA) adopting the supported protocol. After ultrafiltration on 10 000 molecular weight cut off filter (Sartorius AG, Göttingen, Germany) to eliminate protein, nitrate and nitrite was assayed using NO_2^{-}/NO_3^{-} assay (R & D System, Inc, Minneapolis, USA). This assay determines total nitric oxide based on the enzymatic conversion of nitrite by nitrate reductase. The reaction is followed by a colorimetric detection of nitrite as an azo dye product of the Griess reaction (Ding et al, 1988). Separated procedure for nitrite and nitrate assays have been performed. Finally the concentration of endogenous nitrite present in each sample was added to each nitrate concentration and

References

- Barone FC, White RF, Elliott JD, Feuerstein GZ, Ohlstein EH (1995). The endothelin receptor antagonist SB 217242 reduces cerebral focal ischemic brain injury. *J Cardiovasc Pharm* **26**: S404–S407.
- Bonte FJ, Tintner R, Weiner MF, Bigio EH, White CL (1993). Brain blood flow in the dementias: SPECT with histopathologic correlation. *Radiol* **186**: 361-365.
- Ding AH, Nathan C, Stuehr DJ (1988). Release of reactive nitrogen intermediates and reactive oxygen intermediates from mouse peritoneal macrophages. *J Immunol* **141:** 2407–2412.
- Eddleston M, Mucke L (1993). Molecular profile of reactive astrocytes: implication for their role in neurologic disease. *Neurosci* 54: 15–36.
- Ehrenreich H, Lange M, Near KA, Annrser F, Shoeller LA, Schmid R, Winkler PA, Kerl JH, Schmiedek P, Goebel FD (1992). Long term monitoring of immunoreactive endothelin-1 and endothelin-3 in ventricular cerebrospinal fluid, plasma, and 24-h urine of patients with subarachnoid haemorrhage. *Res Exper Med* **192**: 257–258.
- Felgenhauer K, Liappis N, Nekic M (1982). Low molecular solutes and the blood cerebrospinal fluid barrier. *Klin Wschr* **60**: 1385–1392.
- Fujimori A, Yanagisawa M, Saito A, Goto K, Masaki T, Mima T, Takakura K, Shigeno T (1990). Endothelin in plasma and cerebrospinal fluid of patients with subarachnoid haemorrhage. *Lancet* **336**: 633.
- Gard AL, Burrel MR, Pfeiffer SE, Ridge JS, Williams WC (1995). Astroglial control of oligodendrocyte survival mediated by PDGF and leukaemia inhibitory factor-like protein. *Development* **121**: 2187–2197.
- Giaid A, Gibson SJ, Herrero MT, Gentleman S, Lefon S, Yanagisawa M, Masaki T, Ibrahim NB, Roberts GW, Rossi ML, Polak JM (1991). Topographical localisation of Endothelin mRNA and peptide immunoreactivity in neurones of the human brain. *Histochemistry* **95**: 303-314.
- Giovannoni G (1998). Cerebrospinal fluid and serum nitric oxide metabolites in patients with multiple sclerosis. *Multiple Sclerosis* **4**: 27–30.

the results have been expressed as μ mol/l of NO₂⁻ plus NO₃⁻. The sensitivity of the nitrite assay was less than 0.22 μ mol/l while the sensitivity of the nitrate assay was less than 0.54 μ mol/l.

Statistical analysis

The statistical evaluation was performed by Kruskal-Wallis test and Mann Whitney test and the values have been considered significant for $P \leq 0.05$.

Acknowledgements

This work was partially supported by a grant from Ricerca Corrente 1998, given to the Don C Gnocchi Foundation, IRCCS, from the Italian Ministry of Health.

- Goodwin JL, Uemura E, Cunnic JE (1995). Microglial release of nitric oxide by the synergistic action of betaamyloid and IFN-gamma. *Brain Res* **692**: 207–214.
- Harland SP, Kuc RE, Pickard JD, Davenport AP (1995). Characterisation of endothelin receptors in human brain cortex, gliomas, and meningiomas. *J Cardiovasc Pharm* **26**: S408-S411.
- Holzwarth JA, Glaum SR, Miller RJ (1992). Activation of endothelin receptors by sarafotoxin regulates Ca2+ homeostasis in cerebellar astrocytes. *Glia* **5**: 239–250.
- Hooper DC, Ohnishi ST, Kean R, Numagami Y, Dietzschold B, Koprowski H (1995). Local nitric oxide production in viral and autoimmune disease of the central nervous system. *Proc Natl Acad Sci USA* **92**: 5312-5316.
- Ikeda M, Sato I, Matsunaga T, Takahashi M, Yuasa T, Murota S (1995). Cyclic guanosine monophosphate (cGMP), nitrite and nitrate in the cerebrospinal fluid in meningitis, multiple Sclerosis and Guillain-Barre syndrome. *Intern Med* **34**: 734–737.
- Jiang MA, Hoog A, Ma K, Nie JX, Olsson Y, Zhang WW (1993). Endothelin-like immune-reactivity is expressed in reactive astrocytes of the human brain. *Neurorep* **4**: 935–937.
- Kohzuki M, Onodera H, Yasujima M, Itoyama Y, Kanazawa M, Sato T, Abe K (1995). Endothelin receptors in ischemic rat brain and Alzheimer brain. *J Cardiovasc Pharm* **26**: S329–S331.
- Lee ME, de la Monte M, Ng S-C, Bloch KD, Quartermous T (1990). Expression of the potent vasoconstrictor endothelin in the human nervous system. *J Clin Invest* **86**: 141–147.
- Lefvert AK, Link H (1985). IgG production within the central nervous system a critical review of proposed formulae. Ann Neurol 17: 13-20.
- Levin ER (1995). Endothelins. New Engl J Med ${\bf 333:}$ ${\bf 356-363.}$
- Lin WW, Chuang DM (1992). Potentiation by Ca2+ ionophores and inhibition by extracellular KCl of endothelin-induced phosphoinositide turnover in C6 glioma cells. *Neurochem Intern* **21**: 293–301.

- Lindholm D, Castren E, Kiefer R, Zafra F, Thoenen H (1992). Transforming growth factor β 1 in the rat brain, increase after injury and inhibition of astrocytes proliferation. *J Cell Biol* **177**: 395–400.
- Ma KC, Nie XJ, Hoog A, Olsson Y, Zang WW (1994). Reactive astrocytes in viral infections of the human brain express endothelin-like immunoreactivity. J Neurol Sci 126: 184–192.
- MacCumber MW, Ross CA, Snyder SH (1990). Endothelin in brain: receptors, mitogenesis and biosynthesis in glial cells. *Proc Natl Acad Sci USA* 87: 2359-2363.
- Merril JE, Ignarro LJ, Sherman RP, Melink J, Lane TE (1993). Microglial cell cytotoxicity of oligodendrocytes is mediated through nitric oxide. *J Immunol* **151**: 2132–2141.
- Merril JE, Benveniste EN (1996). Cytokines in inflammatory brain lesions: helpful and harmful. *Trends in Neuro Surg* **19**: 331–338.
- O'Brien JT, Eagger S, Syed GM, Sahakian BJ, Levy R (1992). A study of regional cerebral blood flow and cognitive performance in Alzheimer's disease. J Neurol 55: 1182–1187.
- Poser CM, Paty DW, Scheinberg L, McDonald WI, Davis FA, Ebers GC, Johnson KP (1983). New diagnostic criteria for multiple sclerosis: guidelines for research protocols. *Ann Neurol* **13**: 227–231.
- Prineas JW (1985). The neuropathology of multiple sclerosis. In: *Handbook of Clinical Neurology* Koetsier JC (ed). Amsterdam, Elsevier. pp. 213-222.
- Qureshi GA, Baig S, Bednar I, Sodersten P, Forsberg G, Siden A (1995). Increased cerebrospinal fluid concentration of nitrite in Parkinson's disease. *Neuroreport* 6: 1642–1644.

- Ross CA, Snyder SH (1990). Endothelin in brain: receptors, mitogenesis and biosynthesis in glial cells. *Proc Natl Acad Sci USA* 87: 2359-2363.
- Rubaniy GM (1992). Potential physiological and pathological significance of endothelins. *Drugs in the Future* **17**: 915–936.
- Stanimirovic DB, Ball R, Mealing G, Morley P, Durkin JP (1995). The role of intracellular calcium and protein kinase C in endothelin-stimulated proliferation of rat type I astrocytes. *Glia* 15: 119–130.
- Suzuki H, Sato S, Suzuki Y, Takekoshi K, Ishihara N, Shimoda S (1990). Increased endothelin concentration in CSF from patients with subarachnoid haemorrhage. *Acta Neurol Scand* **81**: 553-554.
- Tourtellotte WW, Staugaitis SM, Walsh MJ, Shapshak P, Baumhefner RW, Potvin AR, Syndulko K (1985). The basis of intra-blood-brain-barrier IgG synthesis. *Ann Neurol* **17**: 21–27.
- Wong ML, Rettori V, al-Shekhlee A, Bongiorno PB, Canteros G, McCann SM, Gold PW, Licinio J (1996).
 Inducible nitric oxide synthase gene expression in the brain during systemic inflammation. *Nature Medicine* 2: 581-584.
- Yamada G, Hama Y, Kasuya Y, Masaki T, Goto K (1995). Possible sources of endothelin-1 in damaged rat brain. *J Cardiovasc Pharm* **26**: S486–S490.